Caspase recruitment domain 9, microbiota, and tryptophan metabolism: dangerous liaisons in inflammatory bowel diseases.

CURR OPIN CLIN NUTR METAB CARE. 2017 APR LAMAS B, RICHARD ML, SOKOL H. SORBONNE UNIVERSITY-PIERRE AND MARIE CURIE UNIVERSITY BINSERM

Aim

How caspase recruitment domain 9 (CARD9), one of the numerous IBD susceptibility genes, participate to colitis susceptibility by shaping gut microbiota to produce tryptophan metabolites.

KEY POINTS

- Alteration of intestinal microbiota is observed in IBD.
- CARD9 is one of the identified IBD susceptibility genes.
- CARD9 has a role in shaping the bacterial and fungal gut microbiota and it is required for the production of AhR ligands by the microbiota.
- Impaired ability of the microbiota to catabolize tryptophan into AhR ligands increased sensitivity to colitis by altering the interleukin 22 signaling pathway.
- Impaired microbial production of AhR ligands is observed in patients with IBD and correlates with an IBD-associated SNP within CARD9.

1. IMPACT OF THE CASPASE RECRUITMENT DOMAIN 9 -/- MICROBIOTA IN THE SUSCEPTIBILITY TO COLITIS (colonization of germ free mice with WT microbiota and microbiota from CARD9 -/- mice and then they exposed them to DSS)

- They observed an increased susceptibility to colitis with decreased proliferation and increased apoptosis in intestinal epithelial cell of CARD9 -/- to germfree mice.

- Down-expression of IL22,Reg3g ,and Reg3b gut limited

2. TRYPTOPHAN METABOLISM IS IMPAIRED IN CASPASE RECRUITMENT DOMAIN 9 -/- to GERM-FREE MICE

- They determined the concentration of AhR ligands in the feces of our mice. Production of IAA was impaired in feces of CARD9 -/- to germ-free mice

- Their results showed that impaired tryptophan metabolism in the microbiota of the CARD9 -/- mice lead to defective AhR activation which contributes to the susceptibility of mice to colitis by reduction of interleukin 22 production.

3. IMPAIRED ARYL HYDROCARBON RECEPTOR ACTIVITY AND TRYPTOPHAN METABOLITES IN PATIENTS WITH INFLAMMATORY BOWEL DISEASE

- They analyzed fecal samples from individuals with IBD and healthy study participants for their ability to activate AhR. Impaired AhR activity associated with decreased concentrations of IAA and tryptophan were observed in feces of patients with IBD

- These patients were also genotyped for an IBD-associated single-nucleotide polymorphism (SNP) withinnCARD9 (rs10781499)

- The results suggest a connection between IBD, CARD9, and the ability of the microbiota to produce AhR agonists in humans

Correcting impaired microbiota functions, such as ability to produce AhR ligands, is an attractive strategy in IBD.

A long noncoding RNA signature for ulcerative colitis identifies IFNG-AS1 as an enhancer of inflammation

AMERICAN JOURNAL OF PHYSIOLOGY - GASTROINTESTINAL AND LIVER PHYSIOLOGY 2016 VOL. 311 NO. 3, G446-G457 PADUA ET AL

Long noncoding RNAs are defined as sequences of greater than 200 nucleotides in length. They can express introns and exons, which can be alternatively spliced and generally lack open reading frames for protein translation.

IncRNAs appear to have a diverse set of functions in chromatin remodeling, telomere activity, and subcellular structural organization.

IncRNAs have also been implicated in various steps in the posttranscriptional processing of messenger RNAs, including splicing, editing, transport, translation, and degradation.

The IncRNA IFNG-AS1 was associated with the IBD susceptibility loci SNP rs7134599 and is in close proximity to the inflammatory cytokine IFNG.





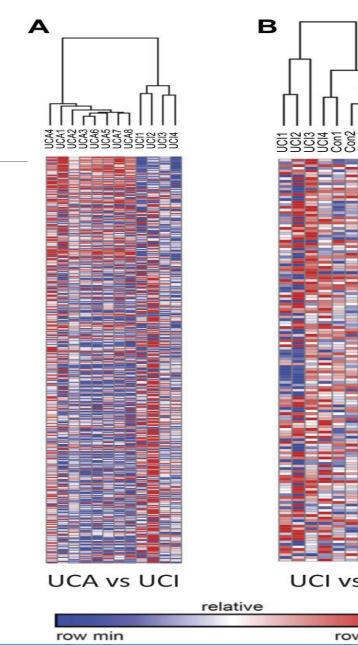


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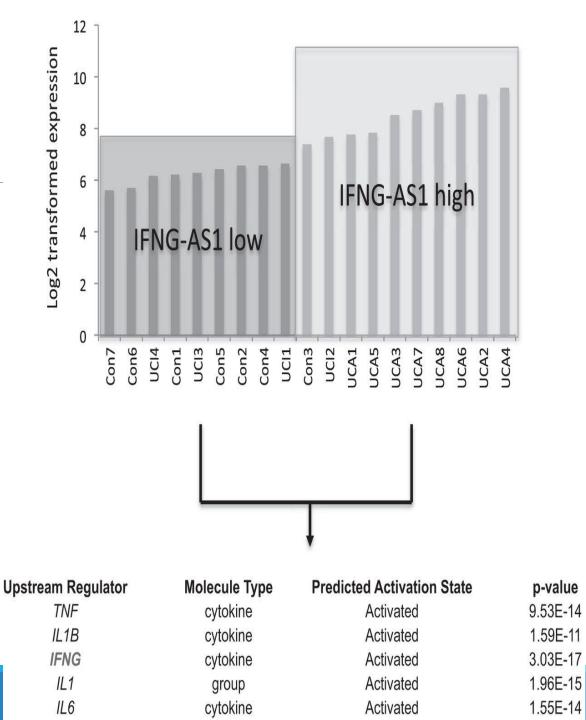
	GeneSymbol	P-value	FC (abs)	Regulation	RNAlength	chrom	strand	txStart	txEnd	
	AC093818.1	2.995E-08	6.5852348	up	474	chr2		173420153	173421135	
	RP11-347C18.3	3.8957E-07	6.0070404	up	709	chr8	+	95962264	95963624	
	AK308561	4.9638E-07	3.3843983	up	640	chr9	4	66454656	66457142	
	AC093818.1	8.4179E-07	6.2669089	up	1397	chr2		173368166	173421324	
	BC015977	1.50809E-06	3.9365962	up	271	chrX		46185713	46187080	
	LOC100505839	1.73017E-06	21.3462294	up	2023	chr10	+	105506536	105515167	
╕╤╡┑╎┍╎┍┾┑	LOC339894	3.09264E-06	6.5386288	up	2016	chr3		156799455	156840791	
NN5 NN2 NN2 NN2 NN2 NN2 NN2 NN2 NN2 NN2	CRNDE	3.11351E-06	4.6952636	up	464	chr16	,	54952965	54957691	
	NOS2P3	3.38483E-06	12.5807175	up	867	chr17	+	20344339	20350085	
	ANKRD36BP2	3.8258E-06	5.0950911	up	391	chr2	+	89065397	89076092	
	RP11-536018.1	5.1115E-06	4.0617397	up	477	chr9	+	13446524	13487510	
	RP11-81H14.2	5.24531E-06	5.9275247	up	417	chr12	4	68825634	68826434	
	BC044655	6.13422E-06	27.8429609	up	2836	chr17	+	20340136	20350557	
	AC133109.1	6.8757E-06	5.5129747	up	1604	chr2	4	109743782	109745386	
	IFNG-AS1	7.05895E-06	5.2737201	up	494	chr12	+	68383308	68628466	
	KCNMB2-IT1	3.0995E-07	2.4886246	down	545	chr3	+	178137021	178175097	
	LOC389023	3.3447E-07	22.8222991	down	744	chr2		115901624	115918920	
	BC043570	2.06129E-06	3.6120905	down	2525	chr15	+	29967193	29971367	
	AC012507.3	4.1393E-06	2.5166143	down	544	chr2	+	231751220	231758789	
	RP11-349K16.1	4.7569E-06	9.2410417	down	552	chr12		128508332	128511711	
	PMS2P3	5.88004E-06	2.0229812	down	1388	chr7		75137068	75157453	
	KRT16P3	7.90624E-06	2.818404	down	681	chr17		20404825	20405600	
	HCG21	8.36127E-06	2.4359782	down	605	chr6		30913755	30922639	
	RP11-10N16.3	1.20406E-05	2.3878804	down	2337	chr1		24620476	24648420	
	RP13-455A7.1	1.2237E-05	2.018203	down	219	chr22	+	48256419	48257811	
	RP11-147/3.1	1.27233E-05	2.0847325	down	469	chr11	+	74624637	74630856	
	RP11-195E11.3	1.32175E-05	2.931462	down	1811	chr9		72784558	72787804	
	PLCD1	1.58876E-05	3.5811373	down	3053	chr3	·	38048986	38066278	
	RP11-627G23.1	1.72506E-05	2.3658954	down	4387	chr11	+	134349192	134375507	
relative	HOXD-AS1	1.74911E-05	2.7729262	down	3819	chr2		177037923	177053686	

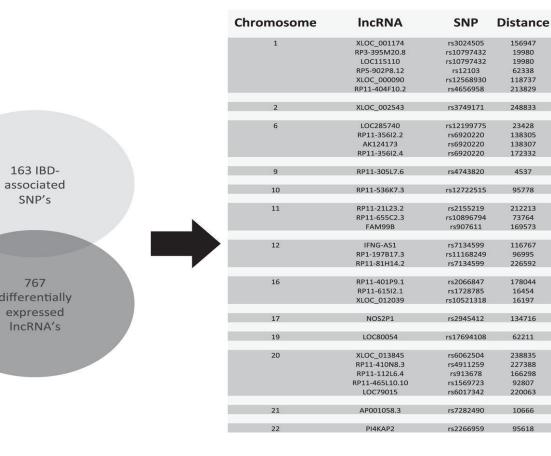




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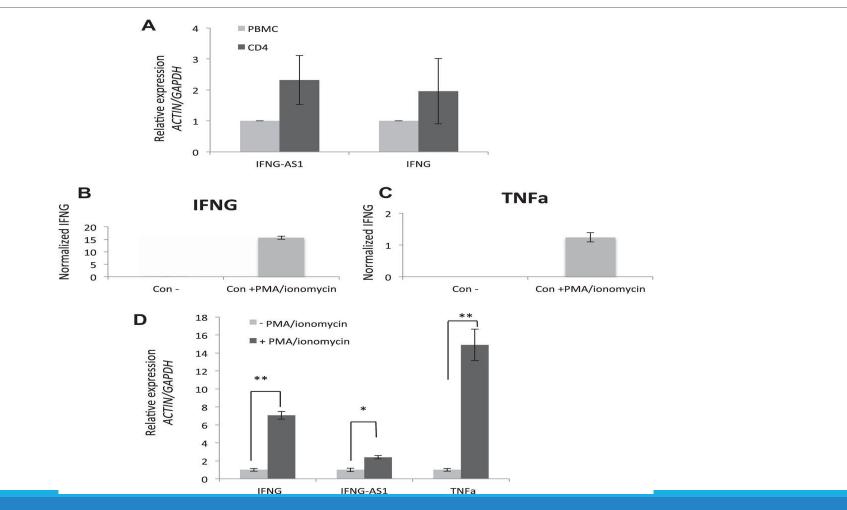




SNP's

767 differentially expressed IncRNA's

qPCR expression of IFNG-AS1 and IFGN among CD4 cells and total PBMC (N = 4). B and C: Jurkat T cells activated with PMA/ionomycin for 6 h and cultured media collected after 24 h show inductions of IFNG and TNF- α protein, respectively. D: Jurkat T cells activated with PMA/ionomycin for 6 h; qPCR analysis of IFNG, IFNG-AS1, and TNF- α . *P value < 0.05, **P value < 0.01; n = 3.



Conclusions

IFNG-AS1 was one of the differentially expressed lncRNAs in UC patients and found to regulate the key inflammatory cytokine, IFNG, in CD4 T cells.

The outhors identified IFNG-AS1 as a novel regulator of IFNG inflammatory responses, suggesting the potential importance of noncoding RNA mechanisms on regulation of inflammatory bowel disease-related inflammatory responses.

Mucosal Expression of Type 2 and Type 17 Immune Response Genes Distinguishes Ulcerative Colitis From Colon-Only Crohn's Disease in Treatment-Naive Pediatric Patients

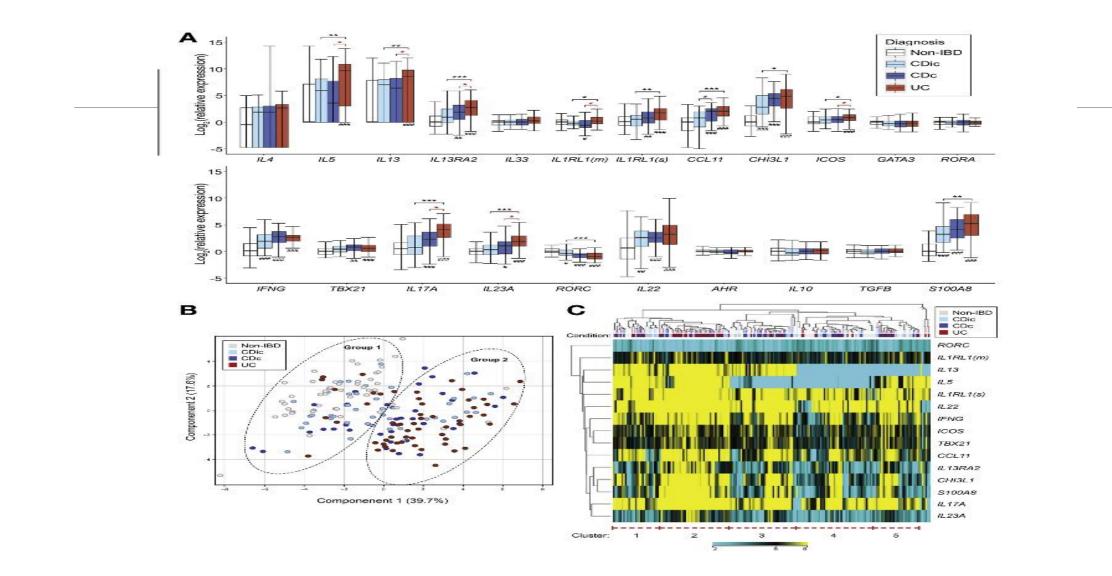
GASTROENTEROLOGY, VOL. 152, ISSUE 6, P1345–1357. MAY 2017 ROSEN ET AL. DIVISION OF GASTROENTEROLOGY, HEPATOLOGY AND NUTRITION, CINCINNATI CHILDREN'S HOSPITAL MEDICAL CENTER, CINCINNATI, OHIO >There is controversy regarding the role of the type 2 immune response in the pathogenesis of ulcerative colitis (UC)

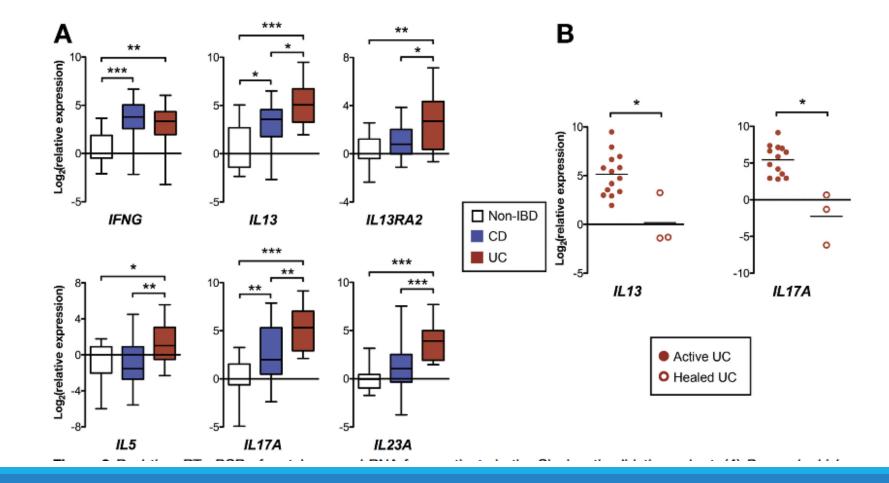
They investigated whether genes associated with a type 2 immune response in the intestinal mucosa are up-regulated in treatment-naive pediatric patients with UC compared with patients with Crohn's disease (CD)-associated colitis or without inflammatory bowel disease (IBD), and whether expression levels are associated with clinical outcomes.

Methodology

They used a real-time reverse-transcription quantitative polymerase chain reaction array to analyze messenger RNA (mRNA) expression patterns in rectal mucosal samples from 138 treatment-naïve pediatric patients with IBD and macroscopic rectal disease, as well as those from 49 children without IBD (controls).

Results were validated in real-time reverse-transcription quantitative polymerase chain reaction analyses of rectal RNA from an independent cohort of 34 pediatric patients with IBD and macroscopic rectal disease and 17controls





Real-time RT-qPCR of rectal mucosal RNA from patients in the Cincinnati validation cohort.

Table 3. Multivariate Logistic Regression for Discriminating UC From CDc						
Gene	OR ^ª	95% CI	P value			

Gene	ORª	95% CI	P value
IL5	1.130	1.032–1.238	.009
IL17A	1.196	0.976–1.467	.085

^aOdds of a diagnosis of UC over CDc per unit increase in Cq value for the listed gene.

The findings support a role for mucosal type 2 inflammatory responses in the early course of pediatric UC.

In treatment-naive pediatric patients, UC is distinguished from Crohn's colitis, and specifically colon-only CD by increased expression of genes associated with type 2 and type 17 immune responses.

Furthermore, an immune gene expression profile marked by increased expression of the type 2 cytokine IL13 is associated with improved clinical outcomes in pediatric UC.

Conclusions

The researchers showed that treatment-naïve pediatric patients with ulcerative colitis exhibit increased mucosal expression of genes associated with type 2 and type 17 immune responses compared to those with colon-only Crohn's disease.

Expression of type 2 and type 17 immune response genes distinguishes ulcerative colitis from colon-only Crohn's disease in treatment-naïve pediatric patients

Genome-wide association study implicates immune activation of multiple integrin genes in inflammatory bowel disease

NATURE GENETICS 49, 256–261 (2017) DE LANGE ET AL.

They performed a genome-wide association study of 25,305 individuals and conducted a metaanalysis with published summary statistics, yielding a total sample size of 59,957 subjects. They identified 25 new susceptibility loci, 3 of which contain integrin genes that encode proteins in pathways that have been identified as important therapeutic targets in inflammatory bowel disease.

The associated variants are correlated with expression changes in response to immune stimulus at two of these genes (ITGA4 and ITGB8) and at previously implicated loci (ITGAL and ICAM1).

In all four cases, the expression-increasing allele also increases disease risk.

They also identified likely causal missense variants in a gene implicated in primary immune deficiency, PLCG2, and a negative regulator of inflammation, SLAMF8.

rs ID	Chr.	Position (bp)	P _{causal}	Effect	Credible set size	Phenotype	P _{meta}	Locus type
rs34687326	1	15,979,9910	1.000	SLAMF8 p.Gly99Ser (missense)	1	CD	1.06 × 10 ⁻⁸	New
rs4845604	1	151,801,680	0.999	RORC (intronic)	1	IBD	7.09×10^{-14}	Known
rs1811711	2	228,670,476	0.914		2	UC	$6.09 imes 10^{-9}$	New
rs56116661	3	188,401,160	0.561	LPP (intronic)	11	CD	5.67×10^{-10}	New
rs11548656	16	81,916,912	0.502	PLCG2 p.His244Arg (missense)	3	IBD	$5.18 imes 10^{-11}$	New
rs1143687	16	81,922,813	0.746	PLCG2 p.Arg268Trp (missense)	5	IBD	3.83 × 10 ⁻⁸	New
rs4821544	22	37,258,503	0.804	NCF4 (intronic)	2	CD	1.76 × 10 ⁻⁸	New

Table 2 Variants fine-manned to 50% probability of being causal in their given signal

Chr., chromosome; CD, Crohn's disease; UC, ulcerative colitis; IBD, inflammatory bowel disease.

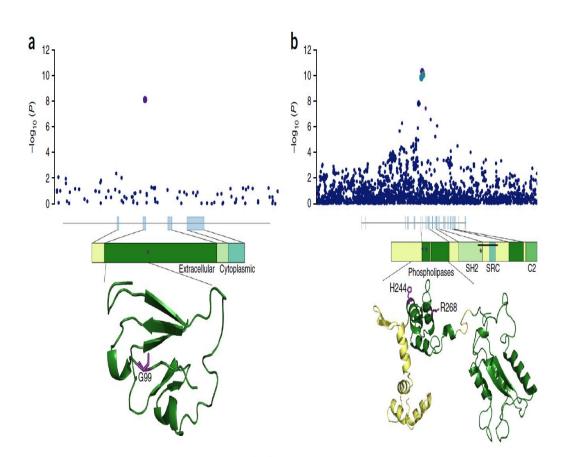


Figure 1 Likely causal missense variants. (**a**,**b**) For *SLAMF8* (**a**) and *PLCG2* (**b**), local association results are plotted with point size corresponding to LD with our lead variant and color corresponding to fine-mapping probability (purple, >50%; intermediate blue, 10–50%; navy blue, <10%). Gene body diagrams and protein domain annotations were taken from Ensembl, and partial predicted crystal structures for both proteins were obtained from the SWISS-MODEL repository.

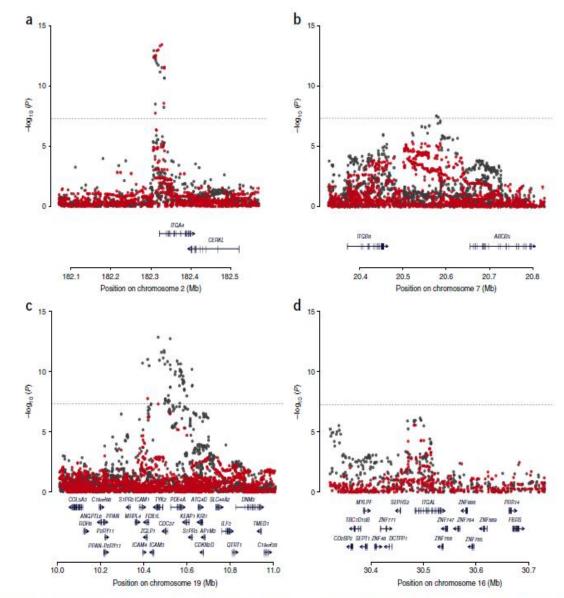


Figure 2 Colocalization of disease associations and stimulus response eQTLs in monocytes. (a–d) The local pattern of disease association for IBD at *ITGA4* (a), *ITGB8* (b) and *ICAM1* (c) or ulcerative colitis at *ITGAL* (d) is shown in gray, and the association of each variant with response to LPS stimulation is shown in red. Evidence of colocalization (probability >70%) is observed for all four loci.

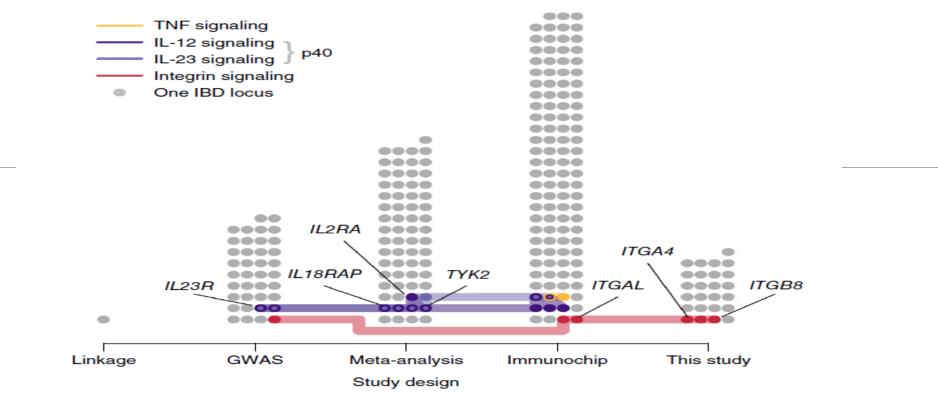


Figure 3 IBD-associated loci containing genes in immune pathways related to classes of approved therapeutics. All IBD susceptibility loci were divided into the studies where they were first identified¹. Loci that contain a gene in one of four signaling pathways related to targets of three classes of approved IBD therapeutics (Online Methods) are highlighted, with those where the pathway gene has been confidently identified as the causal IBD gene labeled. Despite the general pattern of decrease in effect size from left to right, therapeutically relevant associations continue to be found for loci with lower effect sizes.

New associations at common variants continue to identify genes relevant to therapeutic target identification and prioritization.

The Role of the Histone Methyltransferase Enhancer of Zeste Homolog 2 (EZH2) in the Pathobiological Mechanisms Underlying Inflammatory Bowel Disease (IBD).

J BIOL CHEM. 2017 JAN 13;292(2):706-722 SARMENTO ET AL. Regulatory T (Treg) cells expressing the transcription factor FOXP3 play a pivotal role in maintaining immunologic self-tolerance.

It has been shown previously that EZH2 is recruited to the FOXP3 promoter and its targets in Treg cells.

To further address the role for EZH2 in Treg cellular function, they have now generated mice that lack EZH2 specifically in Treg cells (EZH2 Δ/Δ FOXP3+).

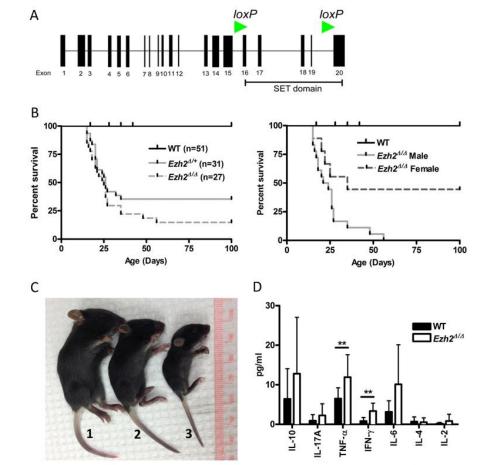


FIGURE 1. **Conditional deletion of the SET domain of EZH2 in FOXP3⁺ Treg cells in mice (EZH2^{\Delta/\Delta} FOXP3⁺ or EZH2^{\Delta/+} FOXP3⁺) results in poor survival.** *A***, exon map of mouse EZH2 indicating the EZH2 catalytic SET domain that was conditionally deleted by flanking LoxP insertion sites (***green arrowheads***).** *B***, survival analysis of EZH2^{\Delta/\Delta} (***n* **= 27) or EZH2^{\Delta/+} (***n* **= 31) mice compared with WT (***n* **= 51) (***left panel***) and mutant mice distinguished by gender (***right panel***). The data are cumulative of over 10 litters, and all offspring are represented.** *C***, representative images depicting the clinical appearance of experimental littermates and the size of FOXP3^{**} EZH2^{D/H} (WT, 1), EZH2^{D/H} FOXP3⁺ (EZH2^{\Delta/+}, 2) and EZH2^{\Delta/+}</sup> FOXP3⁺ (EZH2^{\Delta/+}, 3) pups and lack of ear, eye, and tail inflammation 21 days after birth. D, mean (S.E.) serum cytokine concentrations as measured by multiplex cytokine analysis. Data are from 15 biological**</sup></sup>

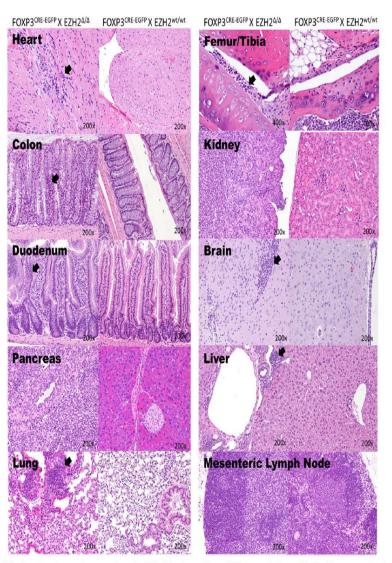
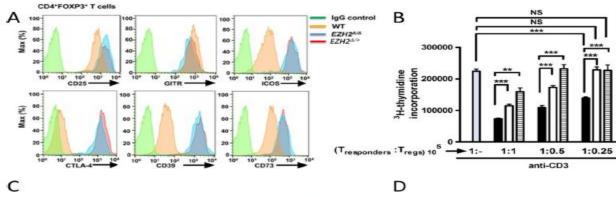


FIGURE 2. EZH2^{Δ/Δ} FOXP3⁺mice develop end organ lymphoid infiltrates, either diffuse, nodular, or both. Shown are the heart, colon, small bowel, pancreas, lung, femoral-tibial joint, kidney, brain, liver, and mesenteric lymph node. Abnormal lymphoid infiltrate is evident in both a diffuse and nodular

EZH2 as a Cofactor for FOXP3 in IBD

Ccr4

Cebpb Pparg Ccl11

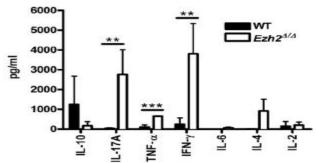


		EZH2KO	EZH2het
Symbol	Thelper categorization	Fold	Fold
II18rap	Th1 Epigen reg gene; cytokine	-11.37	-7.54
lgsf6	Th1 Marker	-2.37	0.00
Irf1	Th1 Marker	0.00	-2.33
Fasl	Th1 Epigen reg gene	0.00	2.02
Socs5	Th1 Marker	2.50	3.60
Tbx21	Th1 Epigen reg gene	2.82	0.00
Tnf	Th1 Marker; cytokine	4.54	4.30
II12b	Th1 Marker; cytokine	5.05	36.73
1118	Th1 Marker; cytokine	6.67	4.59
TIr6	Th1 Marker	6.76	26.71
II18r1	Th1 Epigen reg gene; cytokine	7.70	0.00
ll1r1	Th17 Epigen reg gene; cytokine	-3.27	-4.31
1121	Th17 Epigen reg gene; cytokine	0.00	-2.17
Rorc	Th17 Epigen reg gene; TF	2.07	0.00
II17a	Th17 Epigen reg gene; cytokine	4.24	0.00
II17re	Th17 Epigen reg gene; cytokine	8.49	3.58
115	Th2 Epigen reg gene: cytokine	-5.88	-6.98
Tmed1	Th2 Marker	-2.98	-3.14
Ccl5 (Rar	t Th2 Marker; cytokine	-2.59	-3.90
Ptgdr2	Th2 Marker	0.00	-3.10
Nfatc2	Th2 Marker; TF	0.00	-2.36
Asb2	Th2 Epigen reg gene	0.00	3.41
Nfatc2ip	Th2 Marker; TF	0.00	3.52
81/11	Th2 Epigen reg gene; cytokine	3.21	4.94
II4ra	Th2 Marker; cytokine	3.25	2.36

Th2 Marker; cytokine/recepto

Th2 Marker; TF

Th2 Epigen reg gene Th2 Marker; cytokine



TWE

-:1

Ezh24/+

Ezh2

FIGURE 3. EZH2^{Δ/Δ}FOXP3⁺ lymphocytes transform to a proinflammatory phenotype. A, the expression of cell surface markers in Treg cells (CD4+FOXP3+) measured by flow cytometry. Data are representative of three independent experiments (n = 3 mice/experimental group). B, suppression assay measuring the degree of Treg-mediated suppression of CD4⁺ T responder cells (10⁵) in response to anti-CD3 \pm CD4⁺CD25⁺ Treg titrations (10⁵, 0.5 \times 10⁵ measuring the degree of Treg-mediated suppression of CD4⁺ T responder cells (10⁵) in response to anti-CD3 \pm CD4⁺CD25⁺ Treg titrations (10⁵, 0.5 \times 10⁵ measuring the degree of Treg-mediated suppression of CD4⁺ T responder cells (10⁵) in response to anti-CD3 \pm CD4⁺CD25⁺ Treg titrations (10⁵, 0.5 \times 10⁵ measuring the degree of Treg-mediated suppression of CD4⁺ T responder cells (10⁵) in response to anti-CD3 \pm CD4⁺CD25⁺ Treg titrations (10⁵, 0.5 \times 10⁵ measuring the degree of Treg-mediated suppression of CD4⁺ T responder cells (10⁵) in response to anti-CD3 \pm CD4⁺ CD25⁺ Treg titrations (10⁵, 0.5 \times 10⁵ measuring the degree of Treg-mediated suppression of CD4⁺ T responder cells (10⁵) in response to anti-CD3 \pm CD4⁺ CD25⁺ T reg titrations (10⁵, 0.5 \times 10⁵ measuring the degree of Treg-mediated suppression of CD4⁺ T responder cells (10⁵) in response to anti-CD3 \pm CD4⁺ CD25⁺ T reg titrations (10⁵, 0.5 \times 10⁵ measuring the degree of T reg titrations (10⁵, 0.5 \times 10⁵ measuring the degree of T reg titrations (10⁵, 0.5 \times 10⁵ measuring the degree of T reg titrations (10⁵, 0.5 \times 10⁵ measuring the degree of T reg titrations (10⁵, 0.5 \times 10⁵ measuring the degree of T reg titrations (10⁵, 0.5 \times 10⁵ measuring the degree of T reg titrations (10⁵, 0.5 \times 10⁵ measuring the degree of T reg titrations (10⁵, 0.5 \times 10⁵ measuring the degree of T reg titrations (10⁵, 0.5 \times 10⁵ measuring the degree of T reg titrations (10⁵, 0.5 \times 10⁵ measuring the degree of T reg titrations (10⁵, 0.5 \times 10⁵ measuring the degree of T reg titrations (10⁵, 0.5 \times 10⁵ measuring the degree of T reg titrations (10⁵, 0.5 \times 10⁵ measuring the degree of T reg titrations (10⁵, 0.5 \times 10⁵ measuring the degree of T reg titrations (10⁵, 0.5

2.36 2.77 10.43

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3.31

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6.36

They find that EZH2 deficiency in FOXP3+ T cells results in lethal multiorgan autoimmunity.

They demonstrate that $EZH2\Delta/\Delta FOXP3 + T$ cells lack a regulatory phenotype in vitro and secrete proinflammatory cytokines. Of special interest, $EZH2\Delta/\Delta FOXP3 +$ mice develop spontaneous inflammatory bowel disease.

They assessed the FOXP3 and EZH2 gene networks by RNA sequencing in isolated intestinal CD4+ T cells from patients with Crohn's disease. Gene network analysis demonstrates that these CD4+ T cells display a Th1/Th17-like phenotype with an enrichment of gene targets shared by FOXP3 and EZH2.

Combined, these results suggest that the inflammatory milieu found in Crohn's disease could lead to or result from deregulation of FOXP3/EZH2-enforced T cell gene networks contributing to the underlying intestinal inflammation.